

Supplementary Table S2: Molecular/single-cell scale TNF/TNFR parameters, definitions and values estimated from literature

Parameter	Parameter description	Value*	Reference
$k_{synthMac}$ (#/cell.s)	Full synthesis rate of mTNF for macrophages	10^{-1} -1 (0.21)	(1)
$k_{synthTcell}$ (#/cell.s)	Full synthesis rate of mTNF for T cells	10^{-2} - 10^{-1} (0.021)	(2)
$TNFR1_{mac}$ (#/cell)	TNFR1 density on the surface of macrophages	500-5000 (1100-1900) [†]	(1, 3-5)
$TNFR1_{Tcell}$ (#/cell)	TNFR1 density on the surface of T cells	500-5000 (400-1200) [†]	(1, 3-5)
$TNFR2_{mac}$ (#/cell)	TNFR2 density on the surface of macrophages	500-5000 (400-800) [†]	(1, 3-5)
$TNFR2_{Tcell}$ (#/cell)	TNFR2 density on the surface of T cells	500-5000 (600-800) [†]	(1, 3-5)
D_1 (cm ² /s) [‡]	Diffusion coefficient of sTNF	10^{-8} - 10^{-7} (5.2×10^{-8})	(6, 7)
D_2 (cm ² /s) [‡]	Diffusion coefficient of shed TNF/TNFR2 complex	10^{-8} - 10^{-7} (3.2×10^{-8})	(6, 7)
k_{TACE_Mac} (s ⁻¹)	Rate constant for TNF release by TACE activity on a macrophage	10^{-4} - 10^{-3} (4.4×10^{-4})	(1, 8-10)
k_{TACE_Tcell} (s ⁻¹)	Rate constant for TNF release by TACE activity on a T cell	10^{-5} - 10^{-4} (4.4×10^{-5})	
δ_{TNF} (s ⁻¹)	sTNF degradation rate constant	10^{-4} - 10^{-3} (4.58×10^{-4})	(11)
K_{d1} (M)	Equilibrium dissociation constant of sTNF/TNFR1	10^{-12} - 10^{-10} (1.9×10^{-11})	(3, 12)
K_{d2} (M)	Equilibrium dissociation constant of sTNF/TNFR2	10^{-10} - 10^{-9} (4.2×10^{-10})	(3, 12, 13)
k_{on1} (M ⁻¹ s ⁻¹)	sTNF/TNFR1 association rate constant	10^7 - 10^8 (2.8×10^7)	(12)
k_{on2} (M ⁻¹ s ⁻¹)	sTNF/TNFR2 association rate constant	10^7 - 10^8 (3.5×10^7)	(12)
k_{off1} (s ⁻¹)	sTNF/TNFR1 dissociation rate constant	$k_{on1} \times K_{d1}$	
k_{off2} (s ⁻¹)	sTNF/TNFR2 dissociation rate constant	$k_{on2} \times K_{d2}$	
k_{int1} (s ⁻¹)	TNFR1 internalization rate constant	1.5×10^{-4} - 1.5×10^{-3} (7.7×10^{-4})	(12, 14)
k_{int2} (s ⁻¹)	TNFR2 internalization rate constant	3.9×10^{-4} - 5×10^{-4} (4.6×10^{-4})	(13)
k_{shed} (s ⁻¹)	TNFR2 shedding rate constant	3.9×10^{-4} - 1.5×10^{-3} (5×10^{-4})	(9, 14)
k_{rec1} (s ⁻¹)	TNFR1 recycling rate constant	8.8×10^{-5} - 5.5×10^{-4} (1.8×10^{-5})	(15, 16)
k_{rec2} (s ⁻¹)	TNFR2 recycling rate constant	8.8×10^{-5} - 5.5×10^{-4} (1.8×10^{-5})	(15, 16)
k_{t1} (s ⁻¹)	TNFR1 turn-over rate constant	3×10^{-4} - 5×10^{-4} (3.8×10^{-4})	(15, 16)
k_{t2} (s ⁻¹)	TNFR2 turn-over rate constant	3×10^{-4} - 5×10^{-4} (3.8×10^{-4})	(15, 16)
k_{deg1} (s ⁻¹)	TNFR1 degradation rate constant	10^{-5} - 10^{-4} (5×10^{-5})	(3, 15-17)
k_{deg2} (s ⁻¹)	TNFR2 degradation rate constant	10^{-5} - 10^{-4} (5×10^{-5})	(3, 15-17)
V_{r1_mac} (#/cell.s)	Cell surface TNFR1 synthesis rate constant for macrophages	$k_{t1} \times TNFR1_{mac}$	
V_{r1_Tcell} (#/cell.s)	Cell surface TNFR1 synthesis rate constant for T cells	$k_{t1} \times TNFR1_{Tcell}$	
V_{r2_mac} (#/cell.s)	Cell surface TNFR2 synthesis rate constant for macrophages	$k_{t2} \times TNFR2_{mac}$	
V_{r2_Tcell} (#/cell.s)	Cell surface TNFR2 synthesis rate constant for T cells	$k_{t2} \times TNFR2_{Tcell}$	

* Ranges of parameter values used for sensitivity analysis are indicated out of parentheses. Values in parentheses are used to generate baseline model results.

[†] Baseline model values for TNFR densities on each recruited individual cell was randomly chosen from the range shown in parentheses.

[‡] Diffusion coefficients of the soluble species in granuloma were estimated in line with estimates for diffusible factors of similar molecular weight in tumors (6, 7).

1. Fallahi-Sichani, M., M. A. Schaller, D. E. Kirschner, S. L. Kunkel, and J. J. Linderman. 2010. Identification of key processes that control tumor necrosis factor availability in a tuberculosis granuloma. *PLoS Comput. Biol.* 6: e1000778.
2. Marino, S., D. Sud, H. Plessner, P. L. Lin, J. Chan, J. L. Flynn, and D. E. Kirschner. 2007. Differences in reactivation of tuberculosis induced from anti-TNF treatments are based on bioavailability in granulomatous tissue. *PLoS Comput. Biol.* 3: 1909-1924.
3. Imamura, K., D. Spriggs, and D. Kufe. 1987. Expression of tumor necrosis factor receptors on human monocytes and internalization of receptor bound ligand. *J. Immunol.* 139: 2989-2992.
4. Pocsik, E., R. Mihalik, F. Ali-Osman, and B. B. Aggarwal. 1994. Cell density-dependent regulation of cell surface expression of two types of human tumor necrosis factor receptors and its effect on cellular response. *J. Cell. Biochem.* 54: 453-464.
5. van Riemsdijk-Van Overbeeke, I. C., C. C. Baan, C. J. Knoop, E. H. Loonen, R. Zietse, and W. Weimar. 2001. Quantitative flow cytometry shows activation of the TNF-alpha system but not of the IL-2 system at the single cell level in renal replacement therapy. *Nephrol. Dial. Transplant.* 16: 1430-1435.
6. Nugent, L. J. and R. K. Jain. 1984. Extravascular diffusion in normal and neoplastic tissues. *Cancer Res.* 44: 238-244.
7. Pluen, A., Y. Boucher, S. Ramanujan, T. D. McKee, T. Gohongi, E. di Tomaso, E. B. Brown, Y. Izumi, R. B. Campbell, D. A. Berk, and R. K. Jain. 2001. Role of tumor-host interactions in interstitial diffusion of macromolecules: cranial vs. subcutaneous tumors. *Proc. Natl. Acad. Sci. U. S. A.* 98: 4628-4633.
8. Newton, R. C., K. A. Solomon, M. B. Covington, C. P. Decicco, P. J. Haley, S. M. Friedman, and K. Vaddi. 2001. Biology of TACE inhibition. *Ann. Rheum. Dis.* 60 Suppl 3: iii25-32.
9. Crowe, P. D., B. N. Walter, K. M. Mohler, C. Otten-Evans, R. A. Black, and C. F. Ware. 1995. A metalloprotease inhibitor blocks shedding of the 80-kD TNF receptor and TNF processing in T lymphocytes. *J. Exp. Med.* 181: 1205-1210.
10. Solomon, K. A., M. B. Covington, C. P. DeCicco, and R. C. Newton. 1997. The fate of pro-TNF-alpha following inhibition of metalloprotease-dependent processing to soluble TNF-alpha in human monocytes. *J. Immunol.* 159: 4524-4531.
11. Cheong, R., A. Bergmann, S. L. Werner, J. Regal, A. Hoffmann, and A. Levchenko. 2006. Transient I κ B kinase activity mediates temporal NF-kappaB dynamics in response to a wide range of tumor necrosis factor-alpha doses. *J. Biol. Chem.* 281: 2945-2950.
12. Grell, M., H. Wajant, G. Zimmermann, and P. Scheurich. 1998. The type 1 receptor (CD120a) is the high-affinity receptor for soluble tumor necrosis factor. *Proc. Natl. Acad. Sci. U. S. A.* 95: 570-575.

13. Pennica, D., V. T. Lam, N. K. Mize, R. F. Weber, M. Lewis, B. M. Fendly, M. T. Lipari, and D. V. Goeddel. 1992. Biochemical properties of the 75-kDa tumor necrosis factor receptor. Characterization of ligand binding, internalization, and receptor phosphorylation. *J. Biol. Chem.* 267: 21172-21178.
14. Higuchi, M. and B. B. Aggarwal. 1994. TNF induces internalization of the p60 receptor and shedding of the p80 receptor. *J. Immunol.* 152: 3550-3558.
15. Vuk-Pavlovic, S. and J. S. Kovach. 1989. Recycling of tumor necrosis factor-alpha receptor in MCF-7 cells. *FASEB J.* 3: 2633-2640.
16. Bajzer, Z., A. C. Myers, and S. Vuk-Pavlovic. 1989. Binding, internalization, and intracellular processing of proteins interacting with recycling receptors. A kinetic analysis. *J. Biol. Chem.* 264: 13623-13631.
17. Tsujimoto, M., Y. K. Yip, and J. Vilcek. 1985. Tumor necrosis factor: specific binding and internalization in sensitive and resistant cells. *Proc. Natl. Acad. Sci. U. S. A.* 82: 7626-7630.