After recently re-reading our paper "*Spatial Organization and Recruitment of Non-Specific T Cells May Limit T Cell-Macrophage Interactions Within Mycobacterium tuberculosis Granulomas*", published in January 20, 2021 by Frontiers in Immunology (DOI: 10.3389/fimmu.2020.613638). We identified an issue that we overlooked during our revision of the paper and would like to request a correction be issued for this paper. *Importantly, the issues we identified do not change the interpretation of our data or modify our paper's conclusions*. All the issues relate to Figure 1 of the paper and after we updated the figure the corresponding statistical numbers in Table 2 also were updated (and are even stronger than previous numbers).

Corrections related to Figure 1:
In the original figure, we noted that panels A and C included a stain for CD163 (shown in blue) that was not relevant to our analysis and not included in the Materials and Methods.

- To correct this issue, we CREATED A NEW FIGURE replaced the blue channel in panels A and C (previously CD163) with the DAPI-stained nuclei for these granulomas.
- The channel histograms in Panels A and D were adjusted to improve interpretation of the staining patterns in a way that does not influence the spatial analyses we performed on these granulomas.

We noted that the granuloma shown in panel D had an incorrect channel arrangement that included a stain for neutrophils (shown in red) and the CD11c+ macrophages were depicted in blue. In our analysis of this granuloma, the data we showed in Table 2 represented the interaction between T cells (green) and neutrophils (red) instead of T cells (green) and macrophages (blue in the published figure).

- To correct this issue, we have reorganized the channel coloring to make this granuloma consistent with the other granulomas in panels A-C

- We deleted the neutrophil channel (previously red) and substituted the DAPI channel for this channel so now the channel order includes CD11c+ macrophage (red), CD3+ T cells (green), and nuclei (blue).

- We reanalyzed this granuloma and the updated spatial analysis is indicated in Figure 1 and the corrected numbers have been added to Table 2.

- In the Materials and Methods, we deleted the reference to calprotectin (the neutrophil stain).

Corrections related to Table 2:
Reanalyzing the granuloma in Figure 1, panel D necessitated revisions to Table 2. These revisions include:

- Updating the data for this granuloma, now referred to by an animal-specific identifier that is consistent with the other animals (previously JF13-18, now 20612_29).

- We have updated Table 2 and included it as a separate file.

The changes to the *Materials and Methods* and *Results* sections are indicated in the text below with suggested deletions indicated in red text with a strikethrough (e.g. ~~deletions~~), suggested additions are indicated in bold blue text (e.g. **additions**).

Changes to the text in the Materials and Methods section:

Four randomly selected, formalin fixed paraffin embedded (FFPE) granulomas were derived from **3** cynomolgus macaques (*Macaca fascicularis*), necropsied at approx. **10-11** ~~11, 25-, 25-, and 10-~~ weeks post infection~~, respectively~~ (Figures 1A–D), and were deparaffinized and antigen retrieval was performed as previously indicated (35). Granulomas were stained with cocktails of antibodies including polyclonal rabbit anti-CD3 (Agilent Technologies, Santa Clara, CA), IgG2a mouse anti-CD11c (clone 5D11; Leica Microsystems, Buffalo Grove, IL) ~~and IgG1 mouse anti-calprotectin (also known as S100A9; clone MAC378, ThermoFisher Scientific)~~. Primary antibodies were labeled with fluorochrome-labeled secondaries including anti-isotype (~~IgG1 and~~ IgG2a) specific antibodies (Jackson ImmunoResearch, West Grove, PA). Coverslips were mounted with Prolong Gold with DAPI (ThermoFisher Scientific) and the sections were imaged on an **Olympus Fluoview microscope (Center Valley, PA) or** Nikon e1000 epifluorescence microscope (Nikon Instruments, Melville, NY) with Nikon NIS Elements (Nikon Instruments).

Changes in the results section:

Figure 1 Immunohistochemistry analysis of four non-human primates (NHP) granulomas [shown in Panels **(A-D)**] examining spatial distributions of both T cells and macrophages, and also where they intersect. Four distinct, randomly chosen granulomas images with extracted cell distributions. Column 1 shows the immunohistochemically stained preparation for CD3 (green), CD11c+ (macrophages), and ~~neutrophils~~ **nuclei** (dark blue). White points represent Geographical Information Systems Technology (GIS) analyses of these images revealing cell locations for T cells (Column 2), Macrophages (Column 3), and their intersections (Column 4), as follows. Rows represent four distinct granulomas. The data for the cell numbers in these granulomas are given in Table 2. On average, about ~~8.5~~ **9.75**% (median 8.6%, **StDev is 4.5%**) of T cells interacted with macrophages.

We found that T cell-macrophage interactions occurred for, on average, only about ~~8.5~~**9.75**% of the T cells identified (median: 8.6%, **StDev: 4.5%**), for at least the four granuloma that we examined (See Table 2).

Addition of an author:

The granulomas included in Figure 1, panels B and C were stained and imaged by Nicole Grant, who was unintentionally not included in the authorship of this paper. These images were instrumental in the analyses included in this work and we are requesting that Mrs. Grant be added to the paper as an author in the third author position. Based on the affiliations already included in this paper, we would request she be identified as:

- Nicole L. Grant [5,7]
- Affiliations and addresses: 5-Department of Infectious Diseases and Microbiology, University of Pittsburgh, Pittsburgh, PA, United States. 7-Department of Microbiology and Molecular Genetics and the Center for Vaccine Research, University of Pittsburgh, Pittsburgh, PA, United States